

RESEARCH ARTICLE

Eating increases oxidative damage in a reptile

Michael W. Butler^{1,*}, Thomas J. Lutz¹, H. Bobby Fokidis² and Zachary R. Stahlschmidt^{3,4}

ABSTRACT

While eating has substantial benefits in terms of both nutrient and energy acquisition, there are physiological costs associated with digesting and metabolizing a meal. Frequently, these costs have been documented in the context of energy expenditure while other physiological costs have been relatively unexplored. Here, we tested whether the seemingly innocuous act of eating affects either systemic pro-oxidant (reactive oxygen metabolite, ROM) levels or antioxidant capacity of corn snakes (*Pantherophis guttatus*) by collecting plasma during absorptive (peak increase in metabolic rate due to digestion of a meal) and non-absorptive (baseline) states. When individuals were digesting a meal, there was a minimal increase in antioxidant capacity relative to baseline (4%), but a substantial increase in ROMs (nearly 155%), even when controlling for circulating nutrient levels. We report an oxidative cost of eating that is much greater than that due to long distance flight or mounting an immune response in other taxa. This result demonstrates the importance of investigating non-energetic costs associated with meal processing, and it begs future work to identify the mechanism(s) driving this increase in ROM levels. Because energetic costs associated with eating are taxonomically widespread, identifying the taxonomic breadth of eating-induced ROM increases may provide insights into the interplay between oxidative damage and life history theory.

KEY WORDS: Digestion, Food intake, Hydroperoxides, Prandial state, Reactive oxygen metabolites, Specific dynamic action

INTRODUCTION

Animals rely upon ingesting organic material to provide the nutrients and energy necessary to survive, grow and reproduce, but this process of digesting and absorbing nutrients comes at an energetic cost, typified by specific dynamic action (SDA; Secor, 2009). In brief, SDA incorporates the increase in metabolic rate that is commensurate with the changes in anatomy and physiology necessary to digest and metabolize a meal, such as increasing the size and activity of organs (e.g. small intestines, heart and kidneys; Secor, 2008). This digestion-dependent increase in metabolic rate affects virtually all animals, from mollusks and arthropods to fishes, birds and mammals (Secor, 2009).

The energetic costs of eating are well documented (Secor, 2008, 2009), but digesting a meal may also have non-energetic costs, such as an up-regulation of the immune system (Luoma et al., 2016) or increased oxidative damage. Reactive oxygen species (ROS) are an inadvertent by-product of oxidative phosphorylation in the

mitochondria during normal metabolic processes. Increases in ROS result in damage to organic molecules unless they are neutralized by an organism's antioxidant capabilities (Costantini et al., 2010). Because oxidative damage is associated with an array of biological functions (e.g. ornamentation, hibernation, immune function and reproductive effort; Costantini et al., 2010), it may be a universal constraint in life history evolution (Dowling and Simmons, 2009; Costantini, 2014). Despite this purported importance of oxidative damage, identifying its main physiological drivers has proven challenging. While factors that increase metabolic rate have been linked to an increase in oxidative damage (e.g. immune challenges, van de Crommenacker et al., 2010; exercise, Costantini et al., 2008b), recent work strongly suggests that increased metabolic rate per se is unlikely to drive increases in ROS and, thus, oxidative damage (reviewed in Speakman and Garratt, 2014; Salin et al., 2015).

While an increased metabolic rate is unlikely to directly increase oxidative damage, indirect links between metabolic rate and oxidative damage remain feasible. For example, during periods of increased metabolic rate (e.g. due to eating or exercise), animals mobilize nutrients that are susceptible to oxidation (e.g. triglycerides, Mateos and Bravo, 2007), thus increasing the amount of oxidative damage that is detected in plasma (Sies et al., 2005; Pérez-Rodríguez et al., 2015). Because triglycerides are highly susceptible to free radical attack (Mateos and Bravo, 2007) and many commercial assay kits quantify oxidative damage using markers of lipid peroxidation, eating-induced increases in circulating triglyceride levels may result in the detection of greater amounts of lipid peroxidation (Pérez-Rodríguez et al., 2015). Indeed, work with humans has documented increases in oxidative damage after eating (Sies et al., 2005; Wallace et al., 2010), termed postprandial oxidative stress. However, humans eating three meals per day may spend up to 75% of their lives in an absorptive state (Secor, 2009), making humans a poor model for investigating links between eating and oxidative damage, given that most animals likely spend significant time periods in a non-absorptive state. Thus, eating-induced variation in oxidative damage may have an underappreciated role in evaluating the extent to which oxidative damage informs life history theory.

Here, we used corn snakes (*Pantherophis guttatus*), which can eat meals between 5% and 45% of their body mass (Crocker-Buta and Secor, 2014), to assess how oxidative status changes with absorptive state while controlling for the effects of circulating levels of nutrients (e.g. triglycerides, Pérez-Rodríguez et al., 2015). We chose this species because snakes generally eat relatively large meals and exhibit a robust SDA (Secor, 2009), but the fivefold increase in metabolic rate (i.e. metabolic scope ~5) of corn snakes (Crocker-Buta and Secor, 2014) is in the bottom third of most snake species (Secor, 2009). Thus, while corn snakes eat larger meals and have larger metabolic scopes than many other taxa, corn snakes are not an extreme example within Serpentes. We predicted that if snakes have the ability to up-regulate antioxidant defenses in response to eating-induced oxidative stress, then oxidative damage

¹Department of Biology, Lafayette College, Easton, PA 18042, USA. ²Department of Biology, Rollins College, Winter Park, FL 32789, USA. ³Department of Biology, Georgia Southern University, Statesboro, GA 30460, USA. ⁴Department of Biological Sciences, University of the Pacific, Stockton, CA 95211, USA.

*Author for correspondence (butlermw@lafayette.edu)

 M.W.B., 0000-0002-8390-2960

should change minimally between the non-absorptive and absorptive states. However, if eating results in an increase in oxidative damage beyond the animal's ability to neutralize this oxidation, then oxidative damage should be detectably higher during absorption. By investigating links between eating and oxidative physiology, we evaluate the extent to which postprandial oxidative stress exists in non-human animals, thus informing discussions regarding how oxidative physiology interfaces with life history theory (Dowling and Simmons, 2009; Speakman and Garratt, 2014).

MATERIALS AND METHODS

Husbandry and experimental procedure

We used 32 (14 males, 18 females) captive corn snakes, *Pantherophis guttatus* (Linnaeus 1766), a non-venomous, medium-sized species native to the southeastern USA (Gibbons and Dorcas, 2005). All individuals were 14–16 months old and were the offspring (1st–3rd generation) of wild snakes caught in Beaufort County, SC, USA. We housed snakes individually in translucent plastic enclosures (27×41×15 cm) in a room with a 12 h:12 h light:dark cycle. Enclosures had subsurface heating elements that allowed snakes to thermoregulate along a gradient of temperatures from 24.5 to 33°C, which encompasses the preferred temperature range for corn snakes (Roark and Dorcas, 2000; Stahlschmidt et al., 2015). We allowed snakes *ad libitum* access to water and offered them food (frozen/thawed adult mice that were 15–20% of the snake's body mass) every 2 weeks throughout the 8 week study, even though snakes do not show starvation stress until >112 days without food (McCue, 2008).

We used intra-cardiac blood draws (0.3 ml) to collect blood samples from each snake at two time points, 1 and 7 days post-feeding (dpf), because the metabolic rate of *P. guttatus* peaks (approximately fivefold greater than baseline) at 1 dpf and declines back to pre-feeding levels at approximately 4 dpf when fed meals of 15–20% of their body mass (Crocker-Buta and Secor, 2014). Thus, 1 dpf denotes the absorptive state while 7 dpf denotes the non-absorptive state. Corn snakes exhibit subtle postprandial thermophily (Roark and Dorcas, 2000; Sievert et al., 2013; Z.R.S., unpublished data), meaning that snakes in the absorptive state may have had a higher body temperature (1–3°C higher) and, thus, a metabolic scope >5. To control for order effects, we randomized sampling order (i.e. half the snakes were first sampled during their absorptive state), and samples were separated by at least one full meal cycle (i.e. the duration between sampling for each snake was 2 weeks). To ensure treatment was not nested within blood-collection session, we sampled snakes that were 1 and 7 dpf during each sampling period. We stored blood samples on ice prior to centrifugation at 2350 g for 5 min, and then stored the plasma fraction at –80°C prior to assays (see below). All procedures were approved by the Institutional Animal Care and Use Committee at Georgia Southern University (protocol no. I14004).

Biochemical assays

We quantified the amount of hydroperoxides, which are pro-oxidant intermediates of lipid peroxidation that lead to the production of oxidative damage, with the d-ROMs test (no. MC002, Diacron International, Italy; hereafter d-ROMs). This test detects reactive oxygen metabolites (ROMs) such as hydroperoxides using Fenton's reaction. We followed the directions of the kit, but amended it for smaller sample volumes *sensu* Costantini et al. (2011). In brief, we mixed 4 µl of the sample with 200 µl of kit reagent, incubated it at 37°C for 75 min, and read the absorbance at 505 nm using a Tecan

(Switzerland) Infinite M200Pro plate reader. We calculated the amount of ROMs relative to control blanks and the kit's calibrator, and we report values as mmol l⁻¹ H₂O₂ equivalents, with larger numbers corresponding to a greater amount of ROMs. We also quantified the antioxidant capacity of the plasma using the OXY-Adsorbent test (no. MC435, Diacron International; hereafter OXY), which measures the plasma's ability to oppose the pro-oxidizing action of hypochlorous acid. Also following Costantini et al. (2011), we amended the kit's instructions by combining 5 µl of a 1:100 dilution of plasma with 200 µl of the hypochlorous acid-based oxidizing solution, incubating the plate for 10 min at 37°C, adding 5 µl of the chromagen solution, and then measuring absorbance at 505 nm using a Tecan (Switzerland) Infinite M200Pro plate reader. We calculated antioxidant capacity relative to control blanks and the kit calibrator, and report values in mmol l⁻¹ HClO neutralized, with larger values corresponding to a more robust antioxidant barrier.

To gauge the metabolic state of the subjects, we measured levels of plasma protein and two plasma lipid markers. We quantified plasma protein concentration using the Coomassie Plus (Bradford) Assay (no. 23236, Thermo Scientific, IL, USA) by combining 10 µl of a sample that was diluted 1:100 with ddH₂O with 300 µl reagent, incubating it at room temperature for 10 min, and then measuring the absorbance at 595 nm using a Tecan (Switzerland) Infinite M200Pro plate reader. We then calculated plasma protein concentrations (µg ml⁻¹) using a standard curve of bovine serum albumin (no. 23209, Thermo Scientific). To quantify lipid markers, we used a spectrometric endpoint assay that quantifies both plasma glycerol and triglyceride concentrations, sequentially (Fokidis et al., 2012, 2011; Neuman-Lee et al., 2015). First, we combined 5 µl of a sample with 240 µl of free glycerol reagent (no. F6428, Sigma-Aldrich, MO, USA), incubated it for 10 min at 37°C, and read the absorbance at 540 nm (primary) and 750 nm (secondary) on a Beckman-Coulter AD200 plate reader to obtain the free glycerol concentration in mmol l⁻¹. Immediately after, we added 60 µl of triglyceride reagent (no. T2449, Sigma-Aldrich) and again incubated it for 10 min at 37°C. We read the absorbance at the same settings, and we subtracted this value from the glycerol concentration to provide the plasma levels of formed (or true) triglycerides.

Statistics

We ran the above assays in duplicate, and found that values for all assays were significantly repeatable (Lessells and Boag, 1987) within sample (ROMs: $P < 0.0001$, $R = 0.98$; antioxidant capacity: $P = 0.0014$, $R = 0.63$; protein: $P < 0.0001$, $R = 0.56$; triglycerides: $P = 0.0006$, $R = 0.58$; glycerol: $P < 0.0001$, $R = 0.80$), so we subsequently averaged duplicate samples. We tested for an effect of digestive state, sex and their interaction on ROM, antioxidant capacity, protein, glycerol and triglyceride levels using mixed models with individual as a random effect. We also re-ran the model for ROMs including triglyceride, glycerol or protein level as a covariate to test the possibility that ROM level is proportional to circulating nutrient levels (Pérez-Rodríguez et al., 2015). For one snake (female in the absorptive state), we had enough plasma to run only the d-ROMs and OXY assays, but not to evaluate circulating nutrient levels, and there were several individuals from whom we could not successfully collect plasma in both states (five individuals during the absorptive state, one individual in the non-absorptive state). Lastly, we used Pearson correlations to test whether ROM and antioxidant capacity levels were correlated with each other or with circulating nutrient levels within absorptive or non-absorptive states. All statistics were run using SAS 9.3 (Cary, NC, USA), and

all data are available from the Dryad repository. Residuals from all models were normally distributed, and ROM, antioxidant capacity and glycerol values were log-transformed to meet this assumption of normality. One female had very low levels of antioxidant capacity, which resulted in non-normally distributed residuals for that model. Exclusion of this female resulted in normally distributed residuals, and results were qualitatively similar with or without her exclusion.

RESULTS

There were no differences in antioxidant capacity (both $F_{1,22}<0.26$, both $P>0.62$), ROMs (both $F_{1,24}<1.24$, both $P>0.28$) or plasma protein (both $F_{1,23}<2.42$, both $P>0.13$) due to sex or the interaction of sex and digestive state. However, animals in the absorptive state had increased protein (19.8% increase; $F_{1,23}=65.84$, $P<0.0001$), antioxidant capacity (4.3% increase; $F_{1,22}=5.32$, $P=0.0309$) and ROM (154.9% increase; $F_{1,24}=104.89$, $P<0.0001$; Fig. 1) levels. This effect of digestive state on ROMs remained when including covariates of triglyceride (165.8% increase; $F_{1,22}=118.71$, $P<0.0001$), protein (124.9% increase; $F_{1,22}=37.53$, $P<0.0001$) or glycerol (164.6% increase; $F_{1,22}=107.59$, $P<0.0001$) levels. Males circulated higher levels of triglycerides overall ($F_{1,23}=8.97$, $P=0.0065$), but there was no effect of digestive state or an interaction between sex and digestive state on circulating triglyceride levels (both $F_{1,23}<1.15$, both $P>0.30$), nor did sex, digestive state or their interaction affect circulating glycerol levels (all $F_{1,23}<2.80$, all $P>0.11$).

During the non-absorptive state, protein levels were positively correlated with both ROMs ($r=0.42$, $P=0.0210$) and antioxidant capacity ($r=0.50$, $P=0.0048$), and triglyceride levels were positively correlated with antioxidant capacity ($r=0.39$, $P=0.0320$). During the absorptive state, triglyceride levels positively correlated with glycerol levels ($r=0.43$, $P=0.0244$), and negatively correlated with

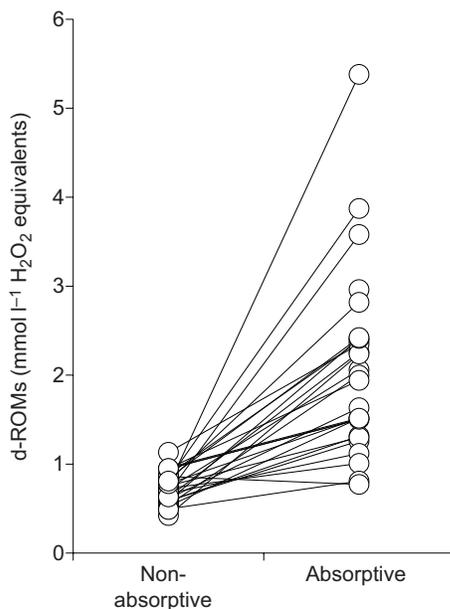


Fig. 1. Oxidative damage in non-absorptive and absorptive states.

Oxidative damage in the plasma was approximately 155% greater ($F_{1,24}=104.89$, $P<0.0001$; mixed model) in the absorptive state ($N=27$) than in the non-absorptive state ($N=31$) in corn snakes. Raw data are plotted, with lines connecting the same individual in both states. Statistics were performed using the log-transformation of d-ROMs values to meet the assumption of normality. Data plotted according to Weissgerber et al. (2015).

ROMs ($r=-0.53$, $P=0.0043$; Fig. 2). All other correlations were not statistically significant (all $r<0.32$, all $P>0.098$).

DISCUSSION

The physiological consequences of eating have primarily been explored via the energetic costs of consuming, digesting and metabolizing a meal, while oxidative costs in non-mammalian systems have been relatively unexplored. Here, we show that digesting a meal has a minimal impact on antioxidant capacity, which increased by 4% – less than differences due to forced locomotion (Costantini et al., 2008b), genetic selection for aggression (Costantini et al., 2008a), or natural variation among populations (Costantini et al., 2009). However, the OXY test does not capture all aspects of antioxidant status of the individual (Sepp et al., 2012), and it is possible that other antioxidant defenses were modulated in ways that we did not detect.

However, despite a minimal increase in antioxidant capacity, we detected a large increase in ROMs (Fig. 1; increases of 1.25 mmol l⁻¹ H₂O₂ equivalents, or 125–166%, depending on statistical model). This magnitude of eating-induced oxidative damage would presumably have important biological consequences, similar to an approximately 130% increase in ROMs due to heat stress (Costantini et al., 2012). Whereas flying 200 km or responding to an immune challenge can increase ROMs by approximately 0.27 mmol l⁻¹ H₂O₂ equivalents (Costantini et al., 2008b) and 0.74 mmol l⁻¹ H₂O₂ equivalents (van de Crommenacker et al., 2010), respectively, we detected an eating-induced increase of over 1.25 mmol l⁻¹ H₂O₂ equivalents. This large increase is likely driven by increases in ROMs, rather than changes in ceruloplasmin or albumin levels, which can interfere with the kit's activity (Kilk et al., 2014; but see Colombini et al., 2016), for two reasons. First, increases in albumin concentration inhibit ROM activity (Kilk et al., 2014); while we quantified circulating protein levels rather than albumin levels per se, we detected greater (not reduced) ROM levels despite increased circulating protein concentration. Second, while ceruloplasmin may create a false positive signal in mammals, the evidence that it affects avian samples is less robust, and its role in non-avian reptile

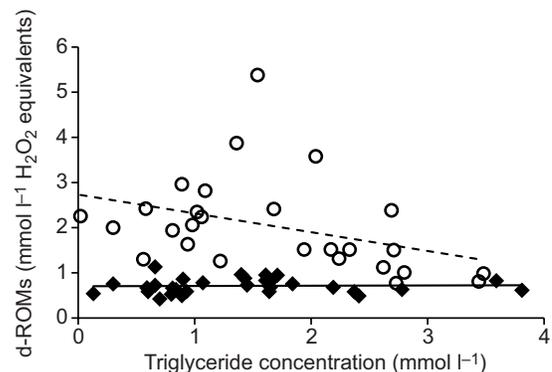


Fig. 2. Oxidative damage versus triglyceride concentration in non-absorptive and absorptive states.

Oxidative damage was not correlated with triglyceride concentration during the non-absorptive state (solid diamonds, solid line; $N=27$; Pearson correlation coefficient=0.06, $P=0.75$), but it was negatively correlated with triglyceride concentration during the absorptive state (open circles, dashed line; $N=30$; Pearson correlation coefficient=-0.53, $P=0.0043$). Raw data are plotted, but statistics were performed using the log-transformation of d-ROMs values. Each snake was sampled at both time points, although limited plasma from several individuals during the absorptive state resulted in a lower sample size during that period.

plasma is unknown (Kilk et al., 2014). Regardless, the repeated-measures nature of our experimental design would require that ceruloplasmin levels would have to change within an individual between digestive states to create the pattern we detected. While data are currently lacking regarding ceruloplasmin levels in reptiles as a function of digestive state, work with chickens has shown that varying levels of food restriction do not affect circulating ceruloplasmin levels (Najafi et al., 2015). Thus, despite methodological concerns regarding the d-ROMs kit, we show that although large meals may have important benefits (e.g. energy acquisition), they can have a greater effect on oxidative damage than traditionally explored stressors. While we predict that the energetic and nutritional benefits of eating will always outweigh the oxidative costs, future work could evaluate the relative magnitude of these costs as a function of meal size, nutrient composition and eating interval.

There are three non-mutually exclusive proximate explanations for increased ROMs during eating. First, increases in ROMs could be due to an increased metabolic rate associated with SDA. While links between metabolic rate and oxidative damage are frequently invoked (Dowling and Simmons, 2009; Speakman and Garratt, 2014), careful consideration of the dynamic state of the mitochondrial membrane strongly suggests that increased metabolic rate per se is unlikely to drive variation in oxidative damage production (Speakman and Garratt, 2014; Salin et al., 2015). For example, higher temperatures increase metabolic rate in corn snakes, but they decrease (not increase) ROMs (Z.R.S., S.S. French and M.W.B., unpublished). Thus, while we did not experimentally test whether an increase in metabolic rate per se causes an increase in oxidative damage, we view this explanation as unlikely based on cellular and molecular (Speakman and Garratt, 2014) and physiological (Salin et al., 2015) evidence.

Second, increases in circulating nutrient levels, and primarily triglycerides, could increase ROM levels through either (a) an increase in the amount of substrates, which would provide a positive feedback loop of oxidizing chain reactions, or (b) a rise in oxidized molecules contained in the food itself, which would then be detected in circulation once they were assimilated (Pérez-Rodríguez et al., 2015). We expected to find such a positive correlation between oxidative damage and nutrient levels based on work with humans (Sies et al., 2005; Wallace et al., 2010) and non-humans (Pérez-Rodríguez et al., 2015). However, ROM levels were positively correlated only with circulating protein levels and only during the non-absorptive state. During the absorptive state, the only correlation between ROMs and circulating nutrient levels was a negative correlation with triglyceride levels. Furthermore, including nutrient levels as covariates in our statistical models as suggested by Pérez-Rodríguez et al. (2015) did not account for the difference in ROMs between absorptive and non-absorptive states. In fact, inclusion of lipid-based markers as covariates yielded a stronger effect of absorptive state on ROM levels.

The third possibility is that, independent of triglyceride levels, the act of digesting a meal resulted in exceptionally high levels of hydroperoxides that were subsequently detected in circulation after being absorbed by the small intestine. While digestion of lipids themselves (e.g. fish oil) does not result in hydroperoxide formation (Larsson et al., 2012), fish oil digested *in vitro* in the presence of hemoglobin does exhibit a marked increase in hydroperoxides (Larsson et al., 2012). Analogously, turkey meat digested *in vitro* shows a substantial increase in hydroperoxide production (Verzelloni et al., 2010). Thus, species consuming moderate amounts of hemoglobin- or iron-containing foods (i.e. meat) may

be more likely to exhibit eating-induced increases in ROMs due to oxidation processes that take place during digestion. Thus, the substantial increase in ROMs that we detected due to eating may be a pattern based more on diet composition than magnitude of SDA.

Regardless of mechanism, we provide two specific insights into the role of oxidative status in life history theory (Dowling and Simmons, 2009). The first ramification is methodological: evaluating the absorptive state of an individual may be critical in interpreting future oxidative physiology studies. Because of the sheer magnitude of the change in ROM levels associated with eating, it is possible that accounting for this variation may reveal biologically important patterns between oxidative status and life history traits that were previously obscured by the digestive state of the animal, particularly for carnivores or species eating iron-rich foods. Second, further work is needed to evaluate the taxonomic breadth of this phenomenon. We performed our experiment with snakes because they exhibit a robust SDA (Secor, 2009). While snakes in general are well known for having some of the largest SDA values (Secor, 2009), almost two-thirds of snakes that have been studied have metabolic scopes greater than 5, demonstrating that corn snakes are not an extreme example within their suborder. Additionally, nearly 25% of anurans and 10% of fishes also have metabolic scopes greater than 5, despite eating meals that are substantially smaller (anurans: 10–15% of body mass; fishes: <10% of body mass) than those of snakes (20–25% of body mass; Secor, 2009). While birds and mammals have SDA-driven metabolic scopes that are substantially lower (e.g. between 1.3 and 2.0; Secor, 2009), the scale of our findings (a greater than twofold increase in ROMs) encourages the exploration of links between digestive state and ROMs even in animals that eat proportionately smaller meals.

Acknowledgements

We thank Lindsey Holcomb for animal husbandry, Tony Mills at the Low Country Institute for the loan of animals, and several anonymous reviewers for comments on previous drafts.

Competing interests

The authors declare no competing or financial interests.

Author contributions

M.W.B. and Z.R.S. were responsible for the conception and design of the experiment and manuscript preparation. T.J.L. and H.B.F. contributed to manuscript revision. M.W.B., T.J.L., H.B.F. and Z.R.S. performed the experiment. M.W.B. carried out statistical analyses. All authors gave final approval for publication.

Funding

Funding was provided by Lafayette College (to M.W.B.), Rollins College (to H.B.F.) and Georgia Southern University (to Z.R.S.).

Data availability

Data are freely available from the Dryad Digital Repository: doi:10.5061/dryad.h020k.

References

- Colombini, F., Carratelli, M. and Alberti, A. (2016). Oxidative stress, d-ROMs test and ceruloplasmin. *Free Radic. Res.* **50**, 447–453.
- Costantini, D. (2014). *Oxidative Stress and Hormesis in Evolutionary Ecology and Physiology: A Marriage Between Mechanistic and Evolutionary Approaches*, 1st edn. Berlin: Springer Heidelberg.
- Costantini, D., Carere, C., Caramaschi, D. and Koolhaas, J. M. (2008a). Aggressive and non-aggressive personalities differ in oxidative status in selected lines of mice (*Mus musculus*). *Biol. Lett.* **4**, 119–122.
- Costantini, D., Dell’Ariccia, G. and Lipp, H.-P. (2008b). Long flights and age affect oxidative status of homing pigeons (*Columba livia*). *J. Exp. Biol.* **211**, 377–381.
- Costantini, D., Dell’Omo, G., De Filippis, S. P., Marquez, C., Snell, H. L., Snell, H. M., Tapia, W., Brambilla, G. and Gentile, G. (2009). Temporal and spatial covariation of gender and oxidative stress in the Galápagos Land Iguana *Conolophus subcristatus*. *Physiol. Biochem. Zool.* **82**, 430–437.

- Costantini, D., Rowe, M., Butler, M. W. and McGraw, K. J. (2010). From molecules to living systems: historical and contemporary issues in oxidative stress and antioxidant ecology. *Funct. Ecol.* **24**, 950–959.
- Costantini, D., Monaghan, P. and Metcalfe, N. B. (2011). Biochemical integration of blood redox state in captive zebra finches (*Taeniopygia guttata*). *J. Exp. Biol.* **214**, 1148–1152.
- Costantini, D., Monaghan, P. and Metcalfe, N. B. (2012). Early life experience primes resistance to oxidative stress. *J. Exp. Biol.* **215**, 2820–2826.
- Crocker-Buta, S. P. and Secor, S. M. (2014). Determinants and repeatability of the specific dynamic response of the corn snake, *Pantherophis guttatus*. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **169**, 60–69.
- Dowling, D. K. and Simmons, L. W. (2009). Reactive oxygen species as universal constraints in life-history evolution. *Proc. R. Soc. B Biol. Sci.* **276**, 1737–1745.
- Fokidis, H. B., Hurlley, L., Rogowski, C., Sweazea, K. and Deviche, P. et al. (2011). Effects of captivity and body condition on plasma corticosterone, locomotor behavior, and plasma metabolites in curve-billed thrashers. *Physiol. Biochem. Zool.* **84**, 595–606.
- Fokidis, H. B., des Rozières, M. B., Sparr, R., Rogowski, C., Sweazea, K. and Deviche, P. (2012). Unpredictable food availability induces metabolic and hormonal changes independent of food intake in a sedentary songbird. *J. Exp. Biol.* **215**, 2920–2930.
- Gibbons, W. and Dorcas, M. (2005). *Snakes of the Southeast*. Athens, GA: University of Georgia Press.
- Kilk, K., Meitern, R., Härmson, O., Soomets, U. and Hörak, P. (2014). Assessment of oxidative stress in serum by d-ROMs test. *Free Radic. Res.* **48**, 883–889.
- Larsson, K., Cavonius, L., Alminger, M. and Undeland, I. (2012). Oxidation of cod liver oil during gastrointestinal in vitro digestion. *J. Agric. Food Chem.* **60**, 7556–7564.
- Lessells, C. M. and Boag, P. T. (1987). Unrepeatable repeatabilities: a common mistake. *Auk* **104**, 116–121.
- Luoma, R. L., Butler, M. W. and Stahlschmidt, Z. R. (2016). Plasticity of immunity in response to eating. *J. Exp. Biol.* **219**, 1965–1968.
- Mateos, R. and Bravo, L. (2007). Chromatographic and electrophoretic methods for the analysis of biomarkers of oxidative damage to macromolecules (DNA, lipids, and proteins). *J. Sep. Sci.* **30**, 175–191.
- McCue, M. D. (2008). Fatty acid analyses may provide insight into the progression of starvation among squamate reptiles. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **151**, 239–246.
- Najafi, P., Zulkifli, I., Soleimani, A. F. and Kashiani, P. (2015). The effect of different degrees of feed restriction on heat shock protein 70, acute phase proteins, and other blood parameters in female broiler breeders. *Poult. Sci.* **94**, 2322–2329.
- Neuman-Lee, L. A., Bobby Fokidis, H., Spence, A. R., Van der Walt, M., Smith, G. D., Durham, S. and French, S. S. (2015). Food restriction and chronic stress alter energy use and affect immunity in an infrequent feeder. *Funct. Ecol.* **29**, 1453–1462.
- Pérez-Rodríguez, L., Romero-Haro, A. A., Sternalski, A., Muriel, J., Mougeot, F., Gil, D. and Alonso-Alvarez, C. (2015). Measuring oxidative stress: the confounding effect of lipid concentration in measures of lipid peroxidation. *Physiol. Biochem. Zool.* **88**, 345–351.
- Roark, A. W. and Dorcas, M. E. (2000). Regional body temperature variation in corn snakes measured using temperature-sensitive passive integrated transponders. *J. Herpetol.* **34**, 481–485.
- Salin, K., Auer, S. K., Rudolf, A. M., Anderson, G. J., Cairns, A. G., Mullen, W., Hartley, R. C., Selman, C. and Metcalfe, N. B. (2015). Individuals with higher metabolic rates have lower levels of reactive oxygen species in vivo. *Biol. Lett.* **11**, 20150538.
- Secor, S. M. (2008). Digestive physiology of the Burmese python: broad regulation of integrated performance. *J. Exp. Biol.* **211**, 3767–3774.
- Secor, S. M. (2009). Specific dynamic action: a review of the postprandial metabolic response. *J. Comp. Physiol. B Biochem. Syst. Environ. Physiol.* **179**, 1–56.
- Sepp, T., Sild, E., Blount, J. D., Männiste, M., Karu, U. and Hörak, P. (2012). Individual consistency and covariation of measures of oxidative status in greenfinches. *Physiol. Biochem. Zool.* **85**, 299–307.
- Sies, H., Stahl, W. and Sevanian, A. (2005). Nutritional, dietary and postprandial oxidative stress. *J. Nutr.* **135**, 969–972.
- Sievert, L. M., Suita, Y. S., Mayes, D. M., Bontrager, L. R., Tuttle, M. and Everly, A. (2013). Specific dynamic action, postprandial thermophily, and the impact of temperature on gastric digestion in the cornsnake. *Pantherophis guttatus*. *Zool. Stud.* **52**, 33.
- Speakman, J. R. and Garratt, M. (2014). Oxidative stress as a cost of reproduction: beyond the simplistic trade-off model. *BioEssays* **36**, 93–106.
- Stahlschmidt, Z. R., Jodrey, A. D. and Luoma, R. L. (2015). Consequences of complex environments: temperature and energy intake interact to influence growth and metabolic rate. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **187**, 1–7.
- van de Crommenacker, J., Horrocks, N. P. C., Versteegh, M. A., Komdeur, J., Tieleman, B. I. and Matson, K. D. (2010). Effects of immune supplementation and immune challenge on oxidative status and physiology in a model bird: implications for ecologists. *J. Exp. Biol.* **213**, 3527–3535.
- Verzelloni, E., Tagliacuzzi, D. and Conte, A. (2010). From balsamic to healthy: traditional balsamic vinegar melanoidins inhibit lipid peroxidation during simulated gastric digestion of meat. *Food Chem. Toxicol.* **48**, 2097–2102.
- Wallace, J. P., Johnson, B., Padilla, J. and Mather, K. (2010). Postprandial lipaemia, oxidative stress and endothelial function: a review. *Int. J. Clin. Pract.* **64**, 389–403.
- Weissgerber, T. L., Milic, N. M., Winham, S. J. and Garovic, V. D. (2015). Beyond bar and line graphs: time for a new data presentation paradigm. *PLoS Biol.* **13**, e1002128.